

Physiological and molecular changes in plants at low temperature

Cold or chilling stress

Fisiologia Molecular do stress

Anabela Bernardes da Silva

Summary

Cold or chilling stress in C4 plants :

- Acclimation to long-term chilling on leaf growth parameters and photosynthesis

Acclimation to long-term chilling in a C4 plant

Effect on leaf growth parameters

Table 1. Effect of low temperature on leaf growth parameters of *P. dilatatum* cv. Raki. Data are means (\pm SE) of 15–16 independent replicates.

	Leaf growth parameters					
	L_{FW} (g)	L_{DW} (g)	% L_{DW}	L_A (cm ²)	L_{FW}/L_A (g FW m ⁻²)	SLA (m ² kg ⁻¹ DW)
Control	0.4736 (\pm 0.0240)	0.0722 (\pm 0.0034)	15.8 (\pm 0.7)	26.29 (\pm 1.21)	179.5 (\pm 1.6)	36.7 (\pm 1.2)
Cold-acclimated	0.3746 (\pm 0.0202)	0.0759 (\pm 0.0051)	20.2 (\pm 0.6)	20.38 (\pm 1.13)	184.2 (\pm 2.1)	27.3 (\pm 9.0)

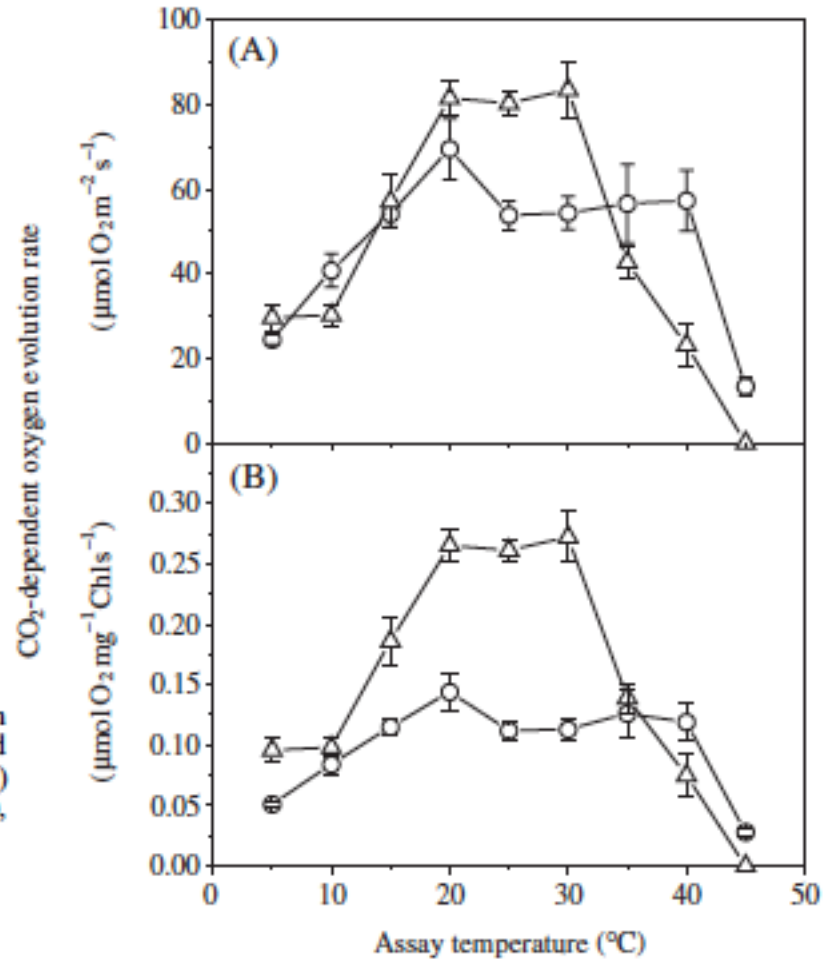
Table 2. Effect of low temperature on photosynthetic pigments leaf content of *P. dilatatum* cv. Raki. Data are means (\pm SE) of 20 independent replicates obtained in two different experiments.

	Photosynthetic pigments content (mg g ⁻¹ FW)					
	Chl <i>a</i>	Chl <i>b</i>	Chl <i>a</i> + <i>b</i>	Carot	Chl <i>a/b</i>	Chl <i>a</i> + <i>b</i> /Carot
Control	2.21 (\pm 0.12)	0.48 (\pm 0.03)	2.69 (\pm 0.14)	0.74 (\pm 0.04)	4.62 (\pm 0.12)	3.63 (\pm 0.04)
Cold-acclimated	1.33 (\pm 0.09)	0.34 (\pm 0.03)	1.67 (\pm 0.11)	0.66 (\pm 0.03)	3.94 (\pm 0.09)	2.48 (\pm 0.07)

Acclimation to long-term chilling in a C4 plant

Effect on photosynthesis

Fig. 2. Temperature response curves of CO₂-dependent oxygen evolution rates in the last fully expanded leaf of control (○) and cold-acclimated (Δ) *P. dilatatum* cv. Raki. Values are means (± SE) of 4–17 independent replicates. (A), A_{CO_2} ($\mu\text{mol O}_2 \text{ m}^{-2} \text{ s}^{-1}$) and (B), A_{CO_2} ($\mu\text{mol O}_2 \text{ mg}^{-1} \text{ Chl s}^{-1}$).



Acclimation to long-term chilling in a C4 plant

Effect on photosynthesis

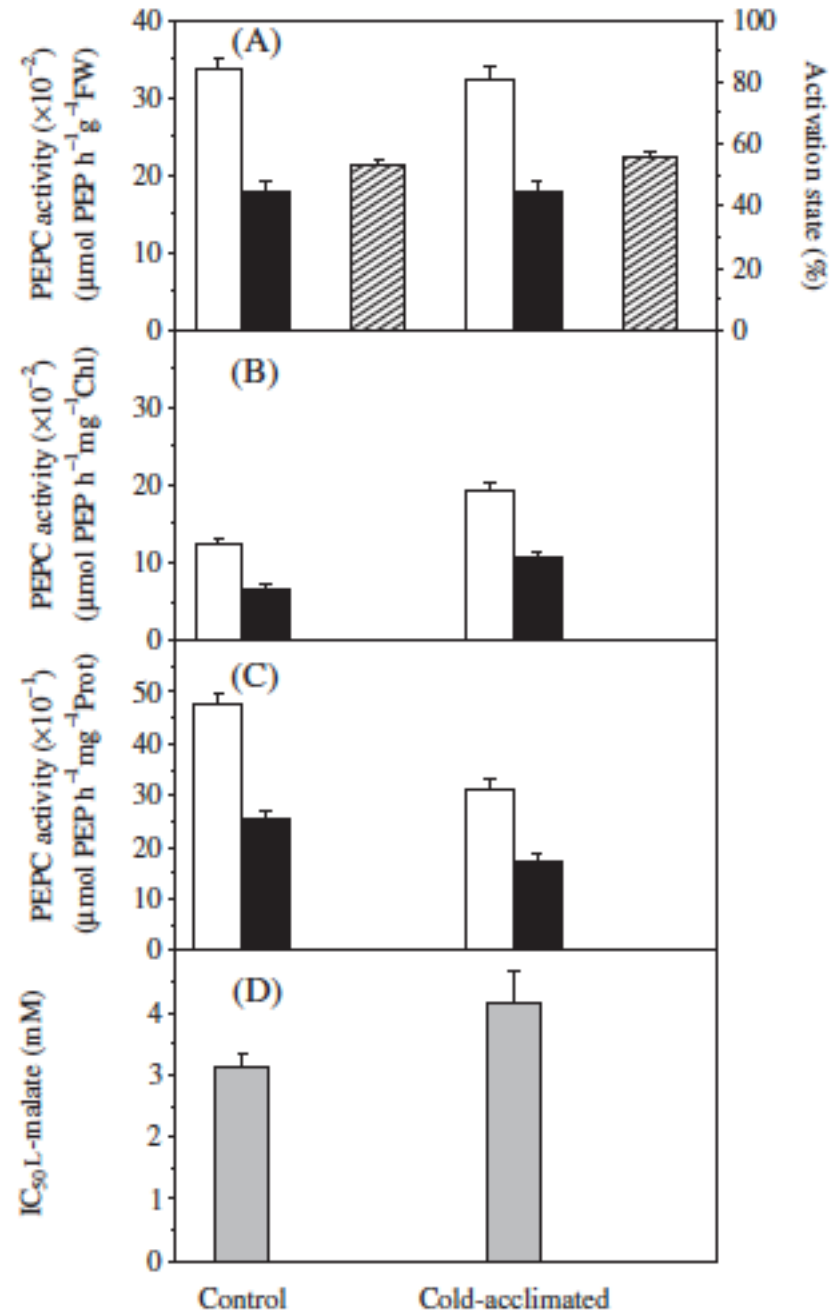
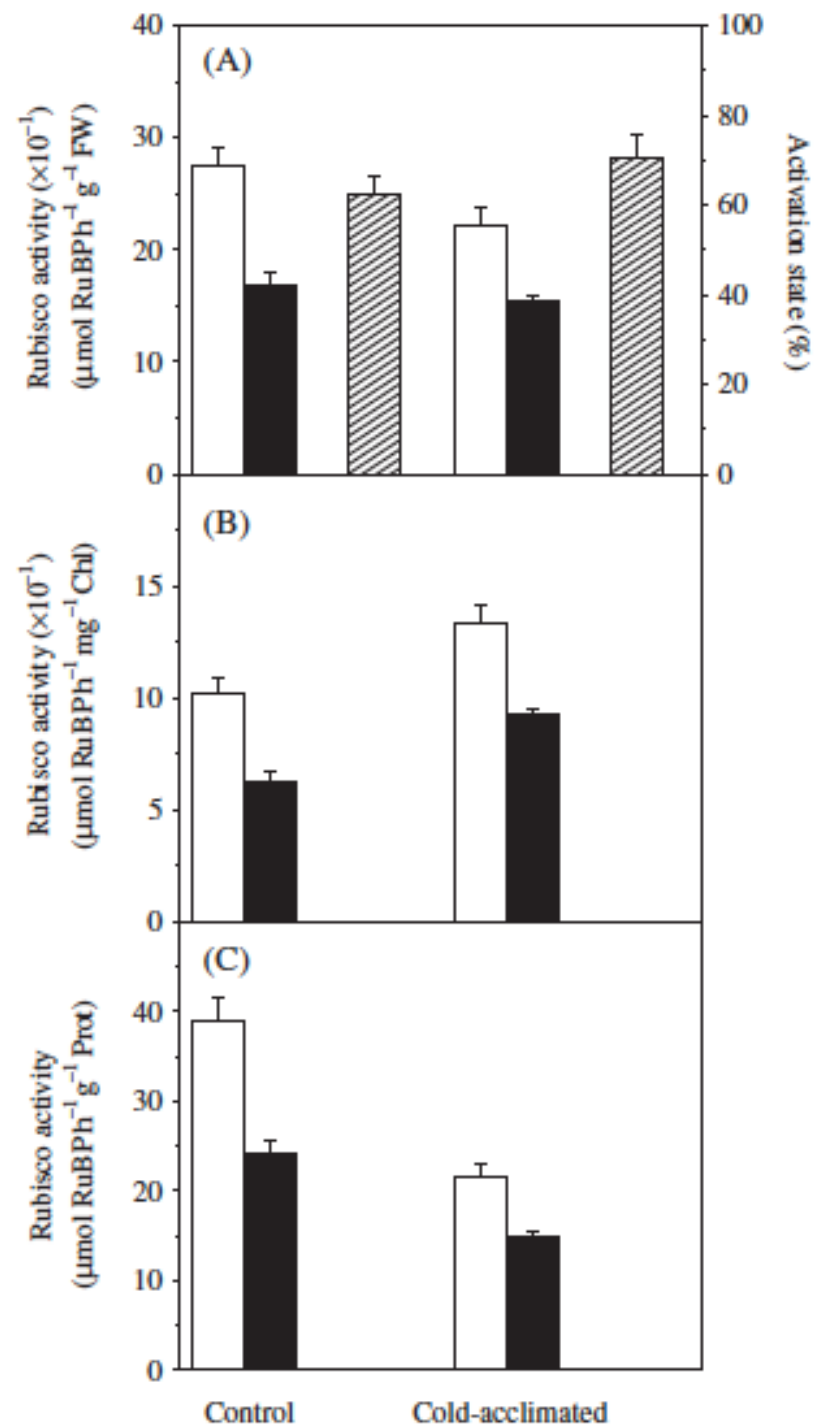


Fig. 6. PEPC activity on a fresh weight basis (A), on a chlorophyll content basis (B) and on a soluble protein content basis (C) and IC₅₀ (L-malate) (D) in the last fully expanded leaf of control and cold-acclimated *P. dilatatum* cv. Raki. Columns are means (+ SE) of six independent replicates. (A)–(C) V_{max} (□), V_{physiol} (■), activation state (▨); (D), IC₅₀ [L-malate] (▨).

Acclimation to long-term chilling in a C4 plant

Effect on photosynthesis

Fig. 7. Rubisco activity [V_{total} (\square), $V_{initial}$ (\blacksquare), activation state (\boxtimes)] on a fresh weight basis (A), on a chlorophyll content basis (B) and on a soluble protein content basis (C) in the last fully expanded leaf of control and cold-acclimated *P. dilatatum* cv. Raki. Columns are means (\pm SE) of six independent replicates.



Acclimation to long-term chilling in a C4 plant

Effect on photosynthesis

Arrhenius eq. $v = A e^{-E_a/RT}$

$\log v = \log A - (E_a/R) (1/T)$

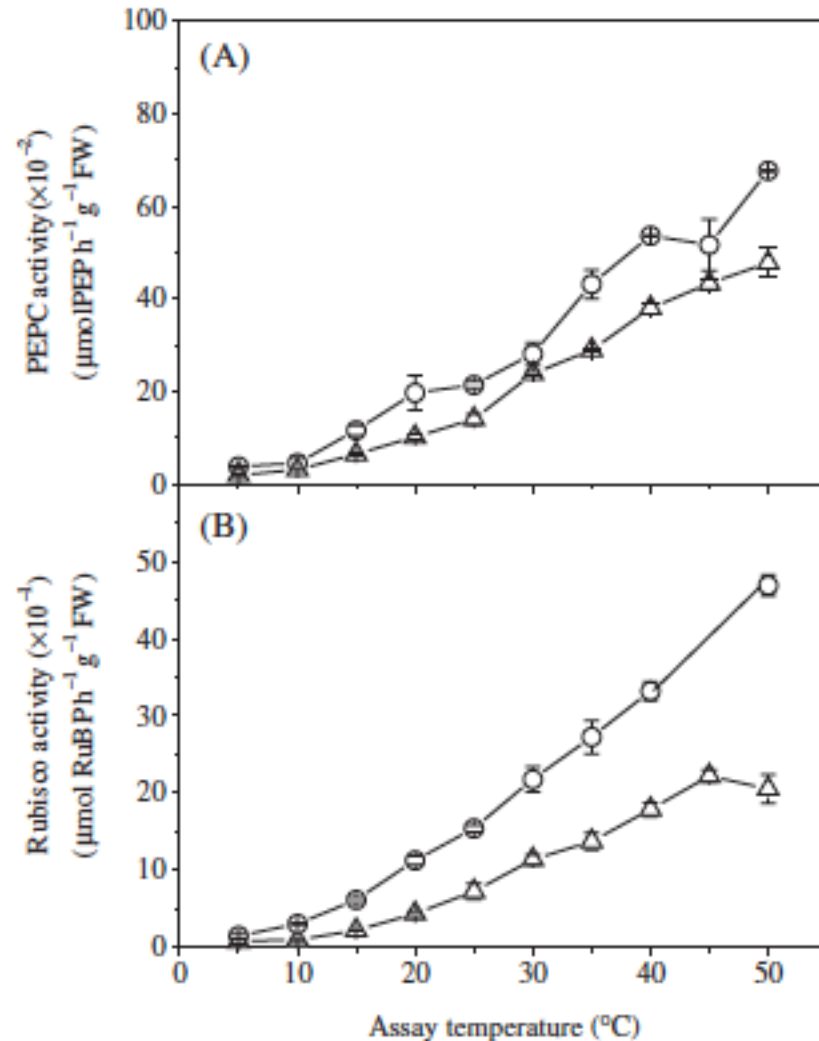


Fig. 8. Temperature curves of PEPC (V_{max}) (A) and Rubisco (V_{total}) (B) activity in the last fully expanded leaf of control (○) and cold-acclimated (Δ) *P. dilatatum* cv. Raki. Values are means (\pm SE) of 2–4 independent replicates.

Acclimation to long-term chilling in C4 plants

Effect on photosynthesis



Figure 1. Emily Heaton next to a stand of giant miscanthus on Caveny Farm, Monticello, Illinois. Each mark on the post equals 1 foot. Photo: John Caveny.

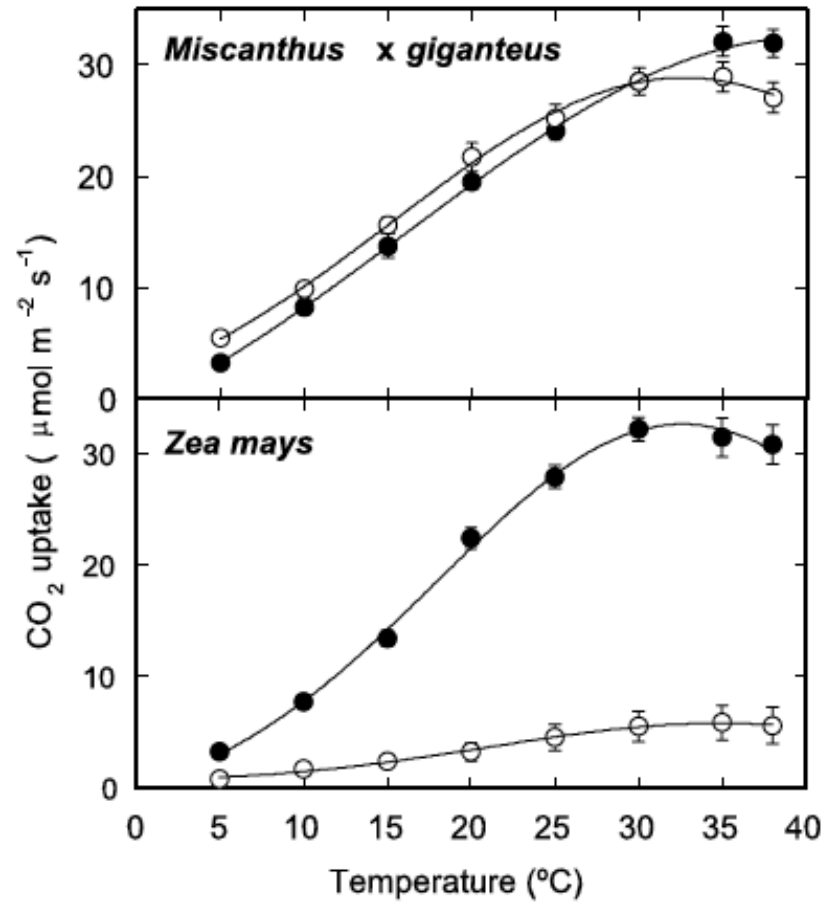


Figure 1. Temperature response of photosynthetic CO₂ uptake per unit leaf area for *M. x giganteus* and maize grown at 25°C/20°C or 14°C/11°C day/night temperatures. Error bars (±1 SE) of the mean ($n = 8-15$) are shown, except when smaller than the symbol size.

Acclimation to long-term chilling in C4 plants

Effect on photosynthesis

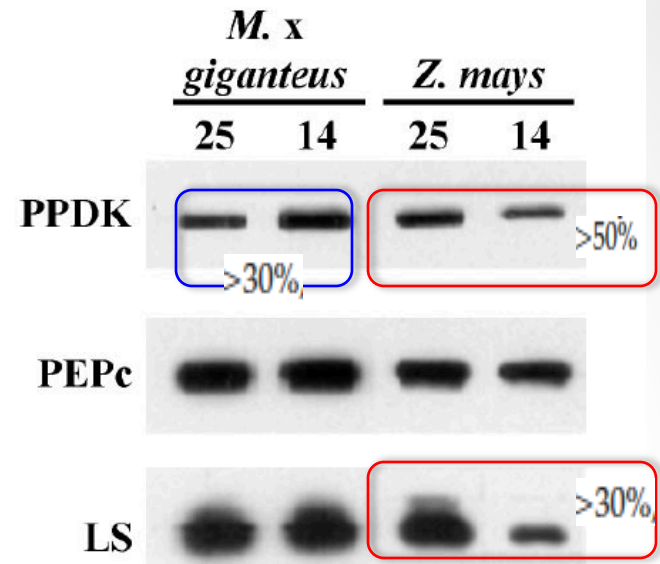


Figure 2. Sample western blot of PPDk, PEPc, and the large subunit of Rubisco (LS) extracted from *M. × giganteus* and maize leaves grown at 25°C/20°C or 14°C/11°C day/night temperatures. Samples were loaded on an equal leaf area basis.

Table I. Protein content

Means and STS of C₄ photosynthetic protein amounts (from western blots) for three replicate leaf samples of *M. × giganteus* and maize plants grown and measured at 25°C/20°C or 14°C/11°C day/night temperatures. Within each replicate, values were standardized to amount of protein in 25°C maize leaves. The bottom row contains values of total soluble protein (grams per square millimeter) extracted from four replicate leaf samples for each species and temperature. Percentage of change is relative to 25°C within each species.

	<i>Miscanthus × giganteus</i>				Maize			
	25°C/14°C	14°C/11°C	Percentage of change	P value	25°C/14°C	14°C/11°C	Percentage of change	P value
PPDK	0.68 (0.172)	0.87 (0.064)	+28%	0.191	1.00 (N/A)	0.43 (0.186)	-57%	0.032
PEPc	0.91 (0.007)	0.90 (0.016)	-1%	0.500	1.00 (N/A)	0.90 (0.053)	-10%	0.032
LS	0.98 (0.016)	1.01 (0.014)	+3%	0.191	1.00 (N/A)	0.61 (0.192)	-39%	0.032
Total	3.73 (0.043)	3.82 (0.035)	+2%	0.131	3.79 (0.037)	3.12 (0.041)	-17%	0.015

Acclimation to long-term chilling in C4 plants

Effect on photosynthesis

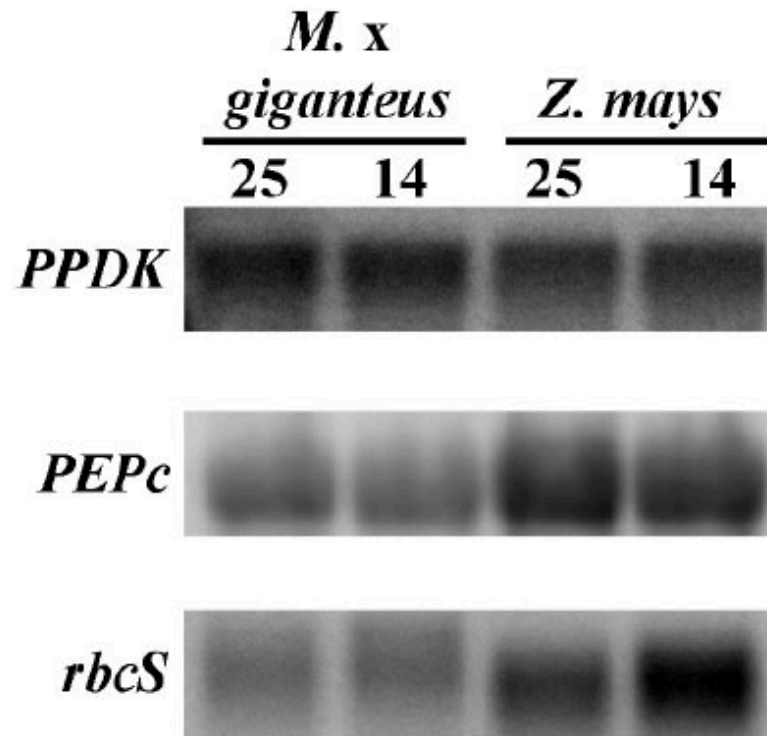


Figure 3. Northern blots of *PPDK*, *PEPc*, and the small subunit of Rubisco (*rbcS*) mRNA from *M. x giganteus* and maize leaves grown at 25°C/20°C or 14°C/11°C day/night temperatures.

Papers to read :

Effects of long-term chilling on growth and photosynthesis of the C₄ gramineae *Paspalum dilatatum*

Ana M. Cavaco^{a,1,*}, Anabela Bernardes da Silva^a and Maria Celeste Arrabaça^a

^aCentro de Engenharia Biológica and Departamento de Biologia Vegetal, Faculdade de Ciências da Universidade de Lisboa, Campo Grande, 1749-016 Lisboa, Portugal

¹Present address: Faculdade de Engenharia dos Recursos Naturais, Universidade do Algarve, Campus de Gambelas, 8000-139 Faro, Portugal

*Corresponding author, e-mail: acavaco@ualg.pt

Received 24 September 2002; revised 14 January 2003

Physiol. Plant. 119, 2003

Cold Tolerance of C₄ photosynthesis in *Miscanthus × giganteus*: Adaptation in Amounts and Sequence of C₄ Photosynthetic Enzymes¹

Shawna L. Naidu, Stephen P. Moose, Abdul K. AL-Shoaibi², Christine A. Raines, and Stephen P. Long*

Department of Crop Sciences, University of Illinois, Urbana, Illinois 61801-4730 (S.L.N., S.P.M., S.P.L.); and Department of Biological Sciences, University of Essex, Colchester, CO4 3SQ, United Kingdom (A.K.A.-S., C.A.R.)

Plant Physiol. Vol. 132, 2003




Tansley review

Advances and challenges in uncovering cold tolerance regulatory mechanisms in plants

Author for correspondence:
Shuhua Yang
Tel: +86 10 62734838
Email: yangshuhua@cau.edu.cn

Received: 8 October 2018
Accepted: 15 January 2019

Yanglin Ding*, Yiting Shi* and Shuhua Yang 

State Key Laboratory of Plant Physiology and Biochemistry, College of Biological Sciences, China Agricultural University, Beijing 100193, China



Epigenetic Control of Plant Cold Responses

Aditya Banerjee¹, Shabir H. Wani^{2,3*} and Aryadeep Roychoudhury^{3*}

¹ Post Graduate Department of Biotechnology, St. Xavier's College-Autonomous, Kolkata, India, ² Mountain Research Centre for Field Crops, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Srinagar, India, ³ Department of Plant Soil and Microbial Sciences, Michigan State University, East Lansing, MI, United States

Keywords: cold stress, DNA methylation, histone modifications, vernalization genes, cold acclimation, crosstalks